

waag society

institute for art, science and technology



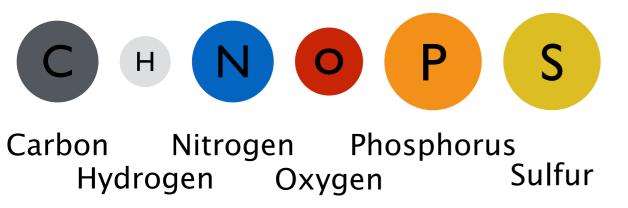
BioHack Kit



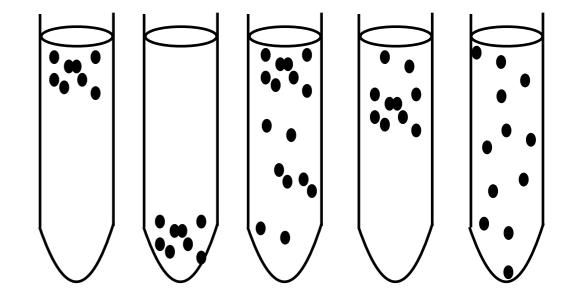


Diversity in growth conditions

Nutrients

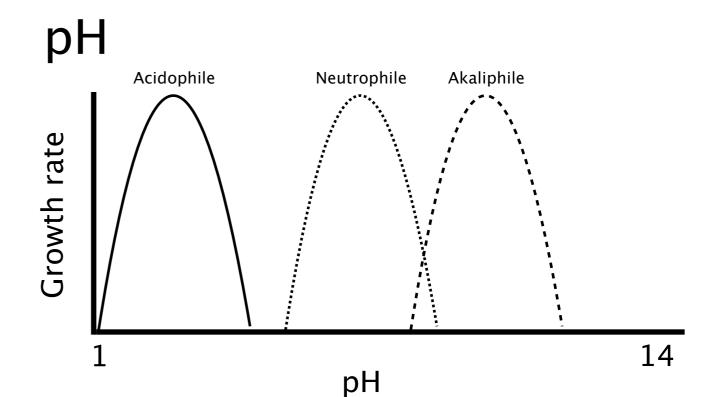


Atmosphere



Temperature







BioFactory canvas



ţţţ input

N

 O_2

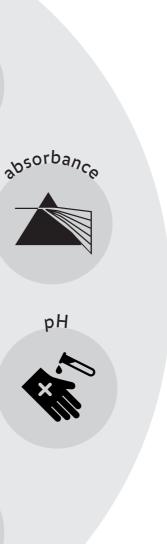
S













observations

day#	
day#	
day#	
day #	
day #	







mass

KG

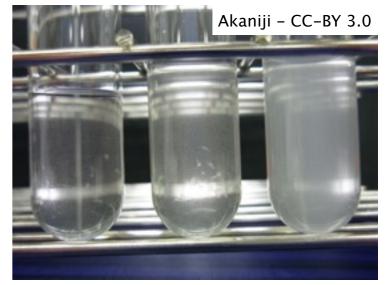


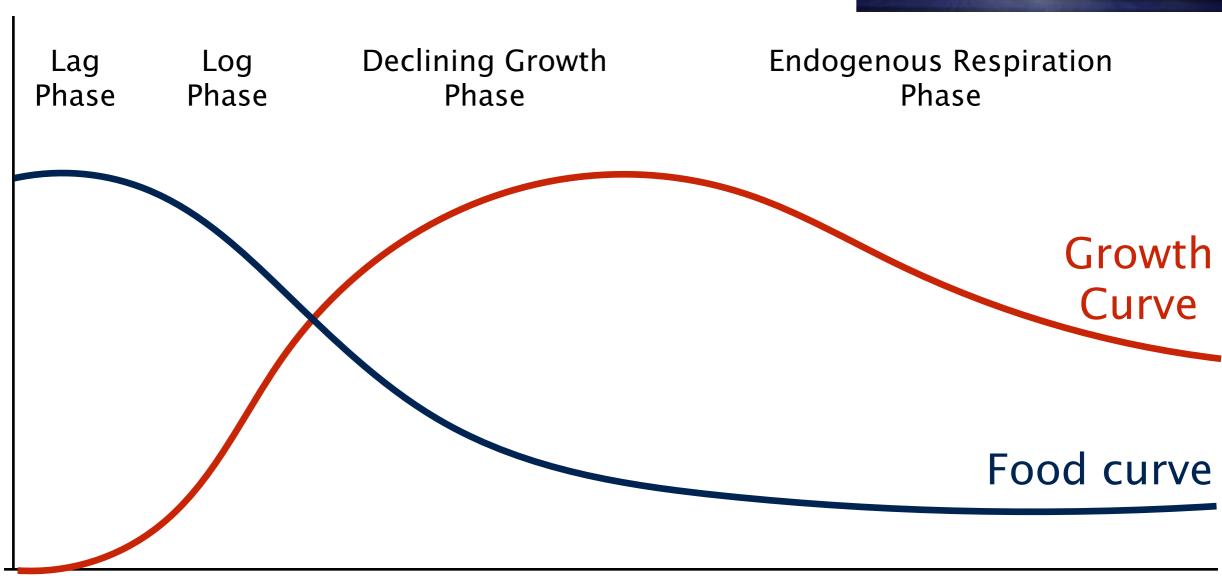
species

time



Bacterial growth curve

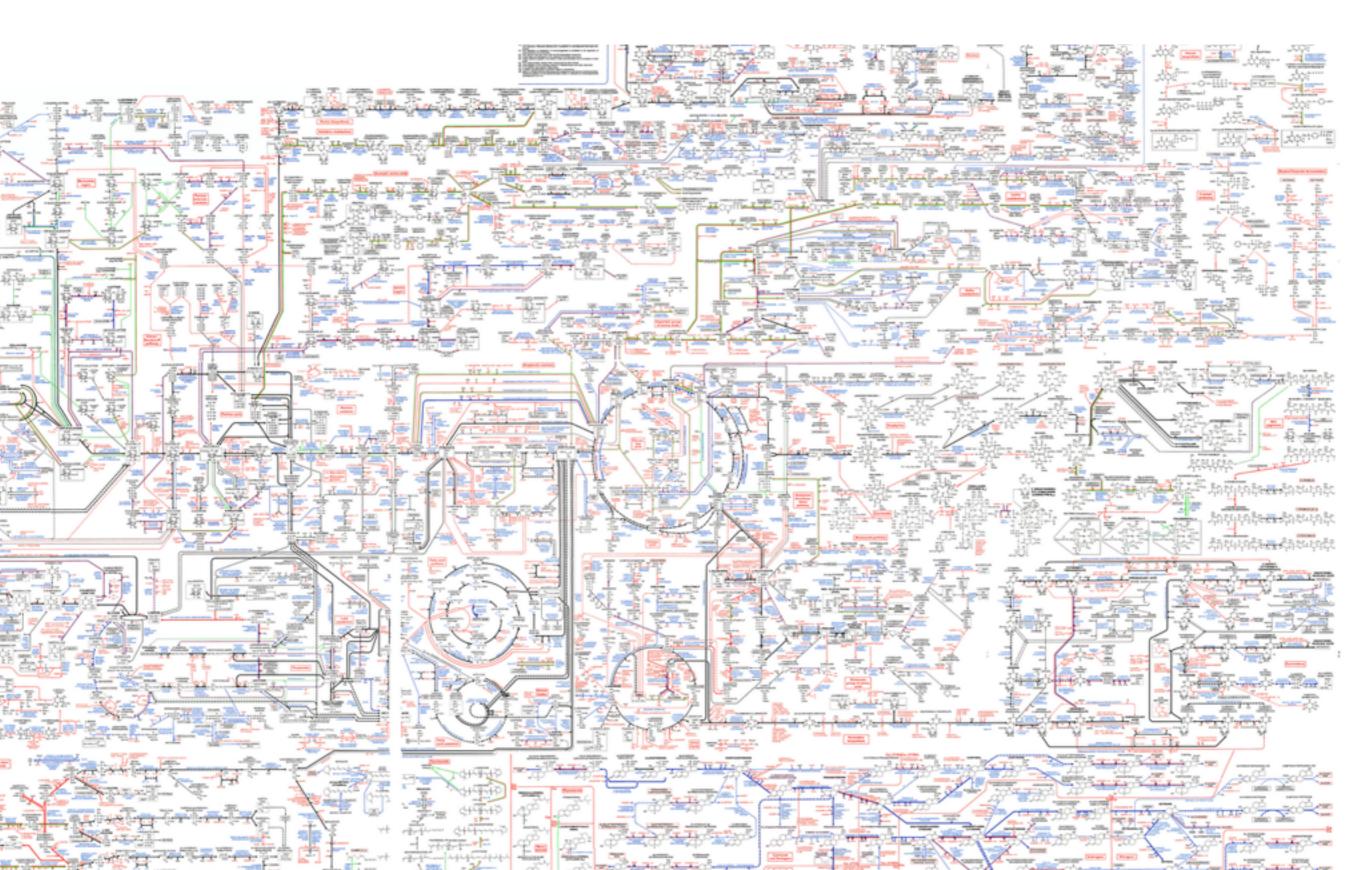




Time

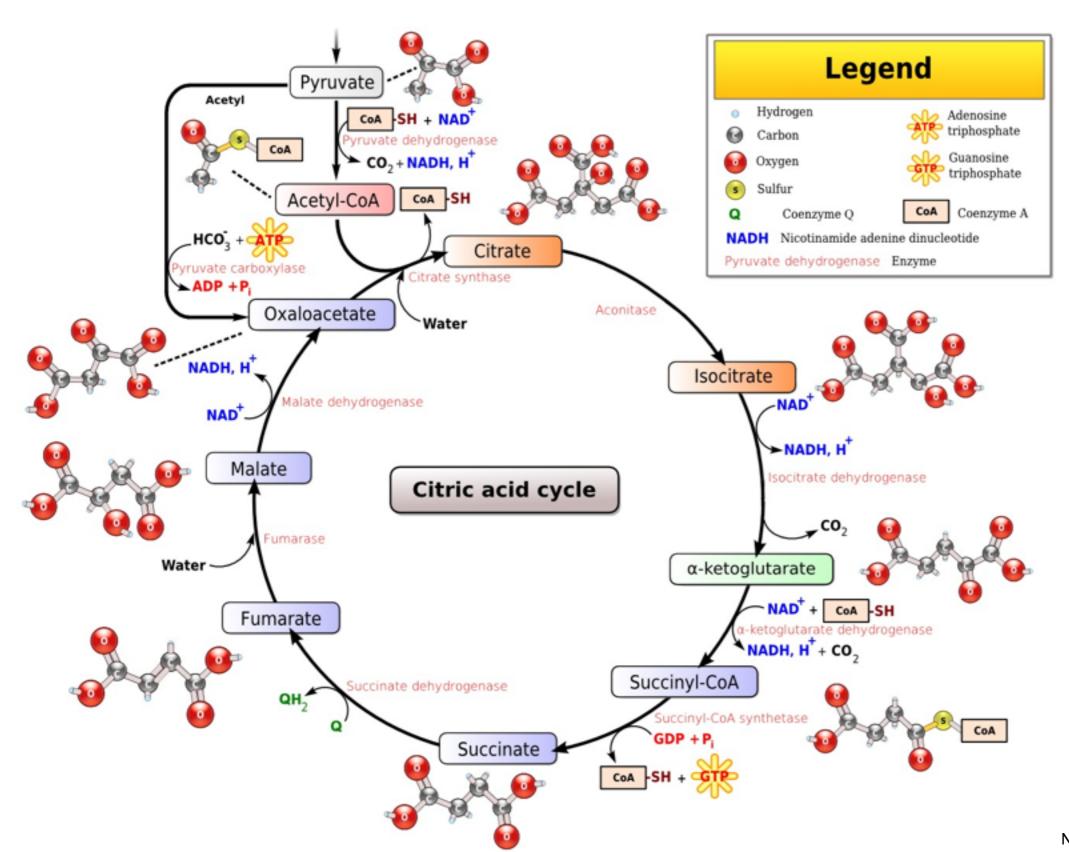


Metabolic Pathways





Citric Acid Cycle





Bioreactor value pyramid

- Antibiotics
- Steroids / hormones
- Vitamins
- Proteins
- Sugars
- Acids

Pharma

Food

Fibers

Fuel

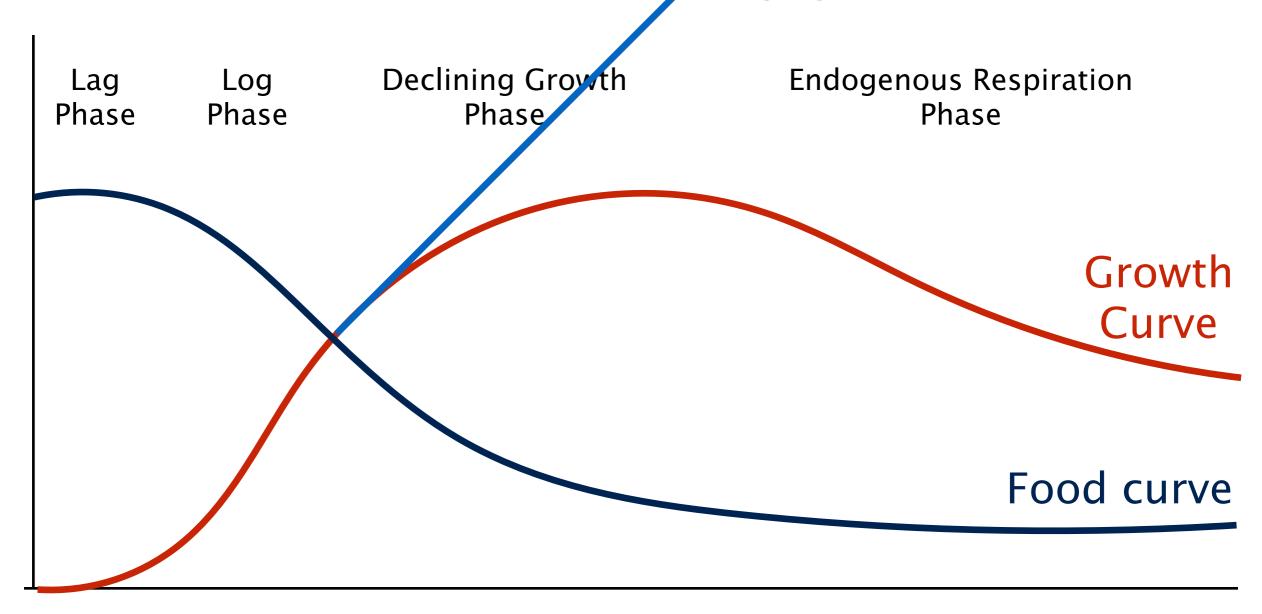
Heat

Volume



Primary products

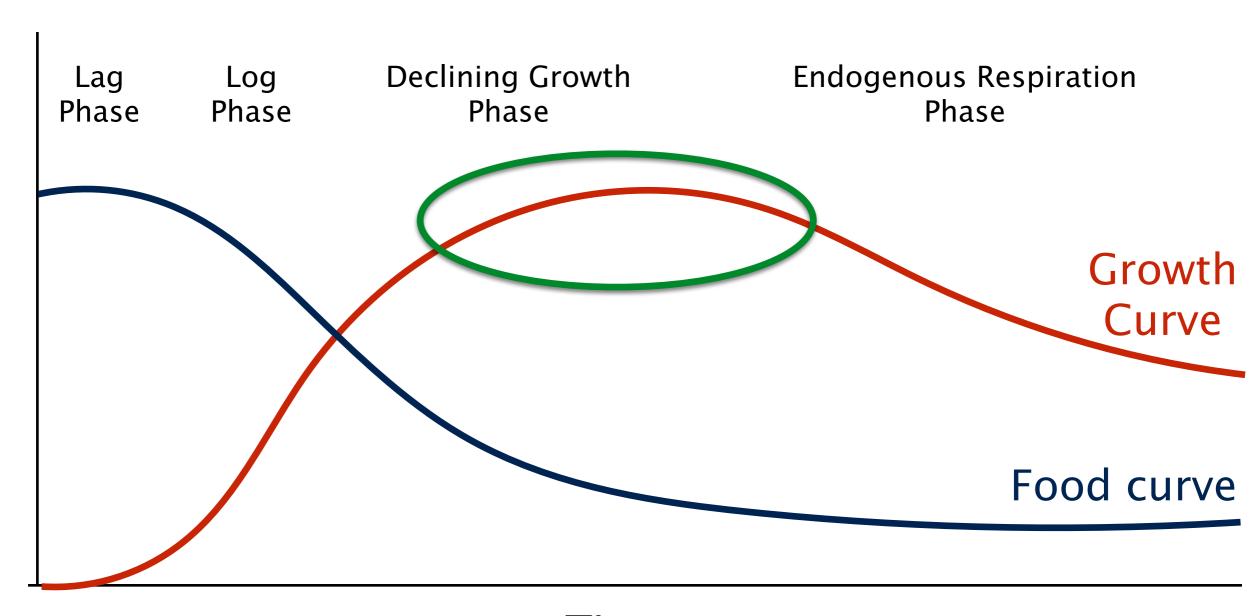
Extend log growth phase



Time



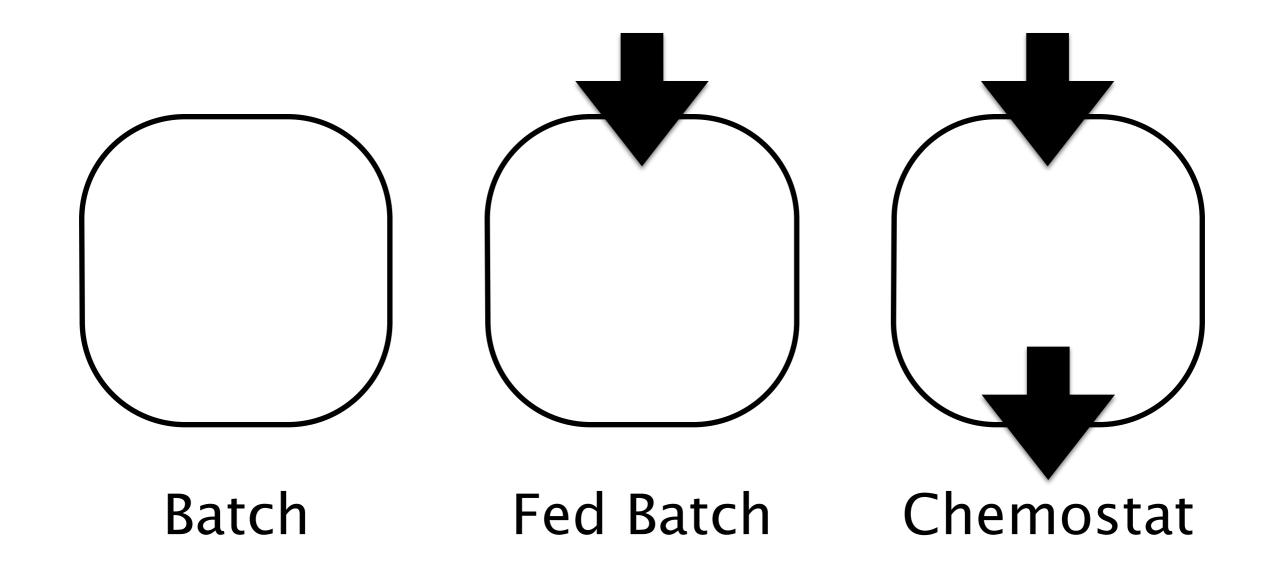
Secondary products



Time



Growth strategies





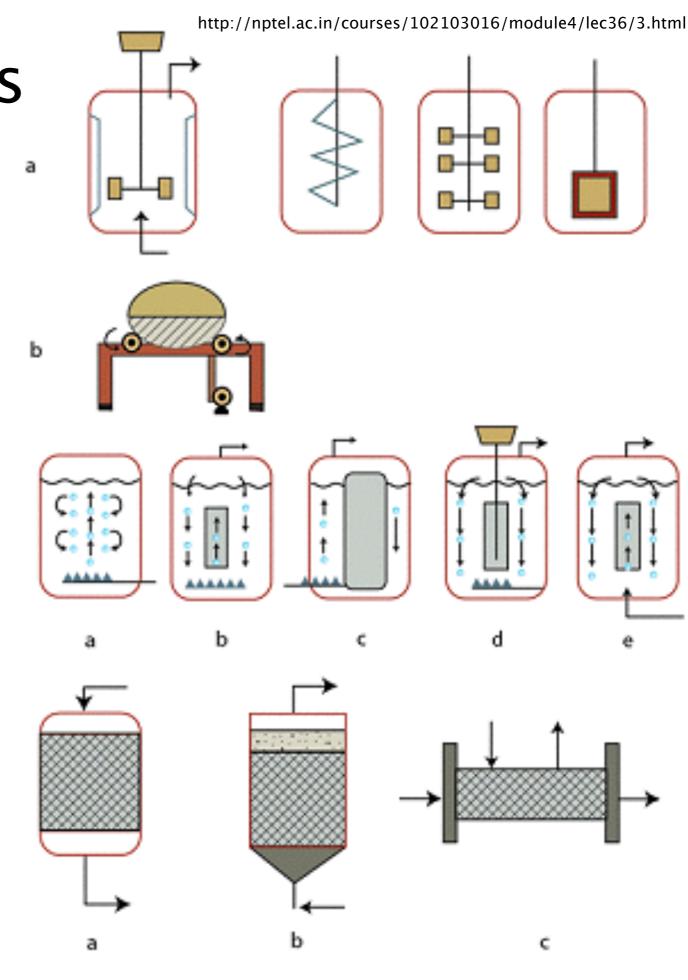
Bioreactor Wishlist

- Simple design
- Many cells per volume
- Uniform distribution
- Simple oxygen supply
- Low energy use



Bioreactor Types

- Stirrer tank
- Air-lift
- Membrane
- Immobilized cells
- Cell culture
- Solid state
- Photobioreactor
- Microbioreactors
- Animals





Bioreactor hacks





Advantages of chemostat

- Measure specific growth speed
- Investigate effect of medium
- Measure & control environmental parameters



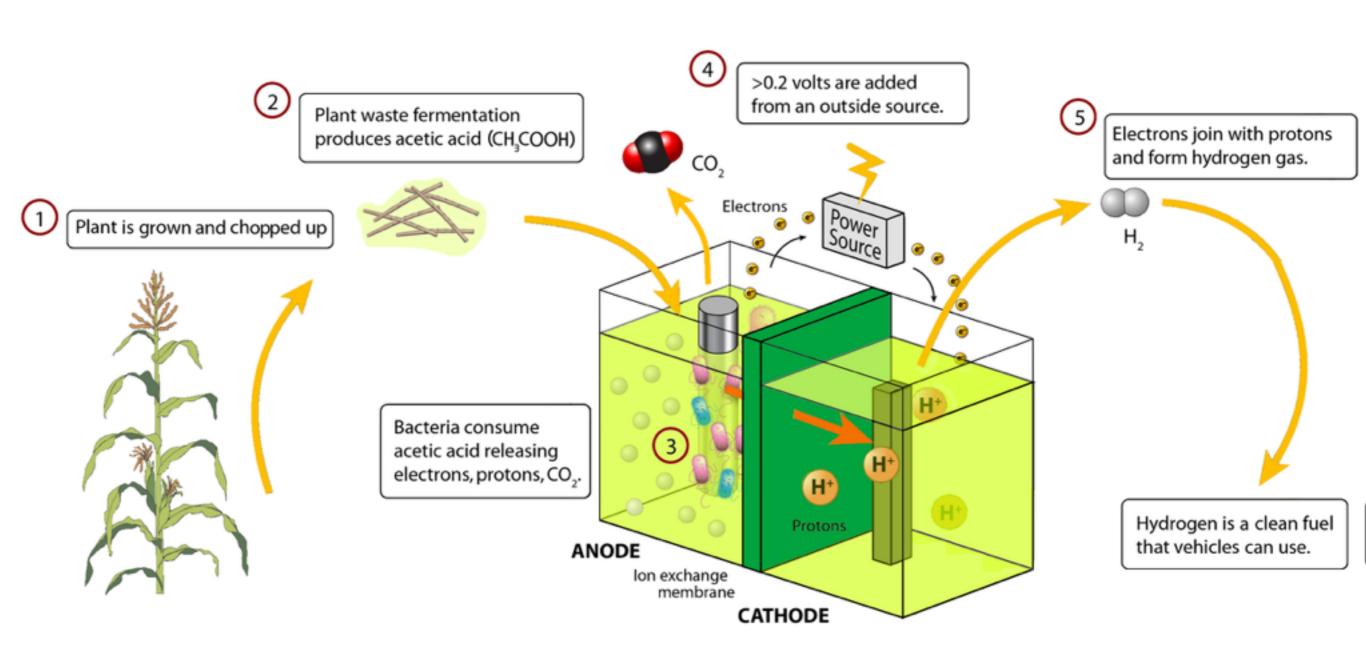


Large scale chemostats





Membrane reactor: Fuel Cells



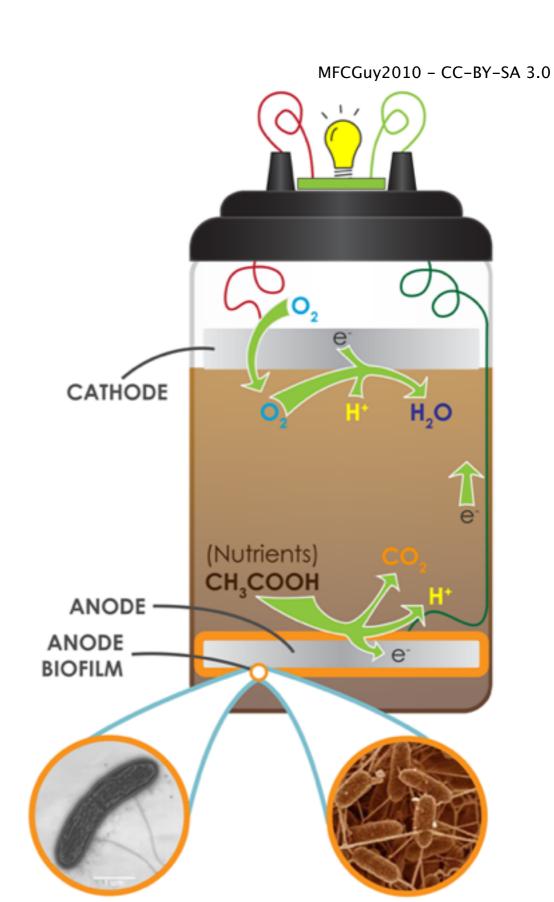
MICROBIAL ELECTROLYSIS CELL

PublicDomain



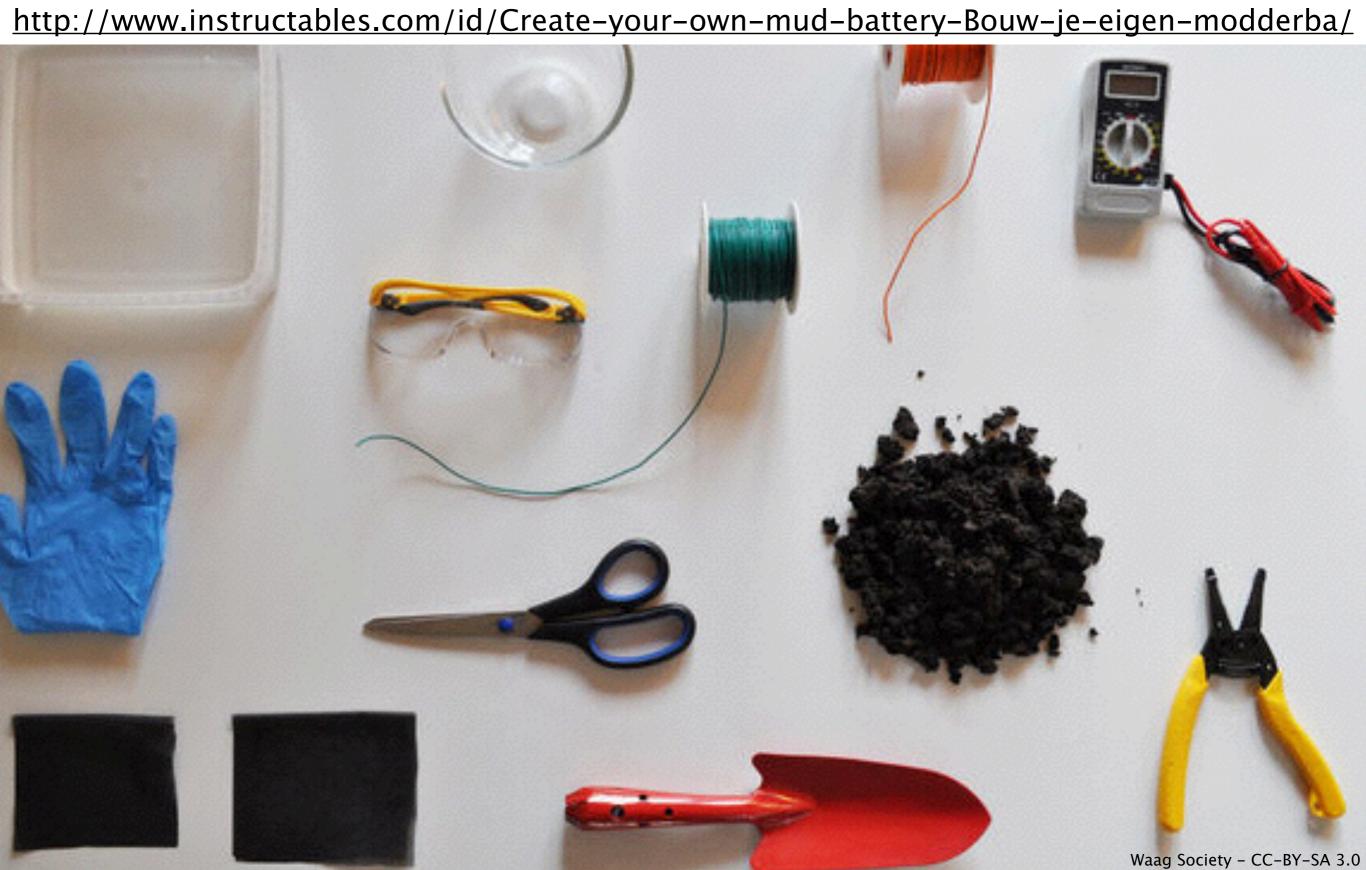
Soil based fuel cells







Bio mud battery





100 m3 reactor

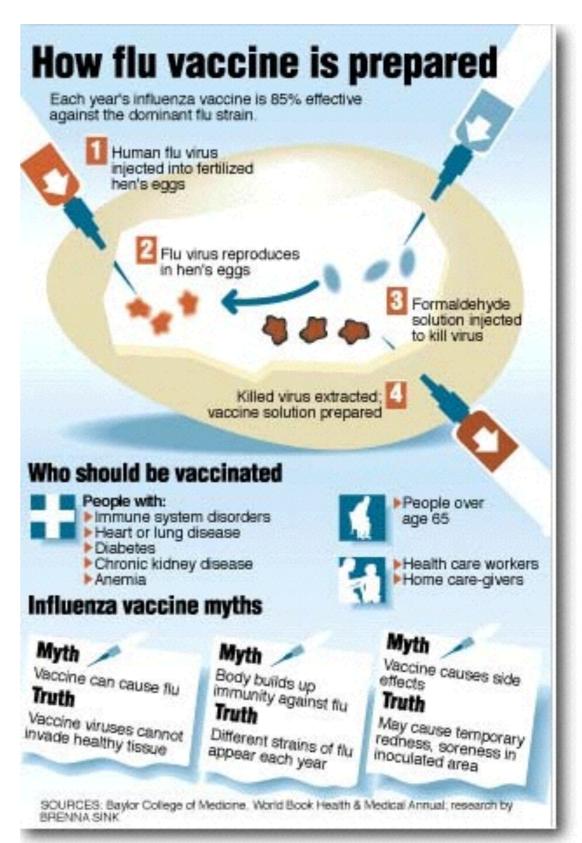
- Yeast production
- Yield = product / substrate
- Fermentation: 2 ATP per sugar
- Respiration: 16 ATP per sugar
- Even at full aeration risk of low yield: Respirofermentive metabolism





Flu vaccine production in eggs







Transport Phenomena

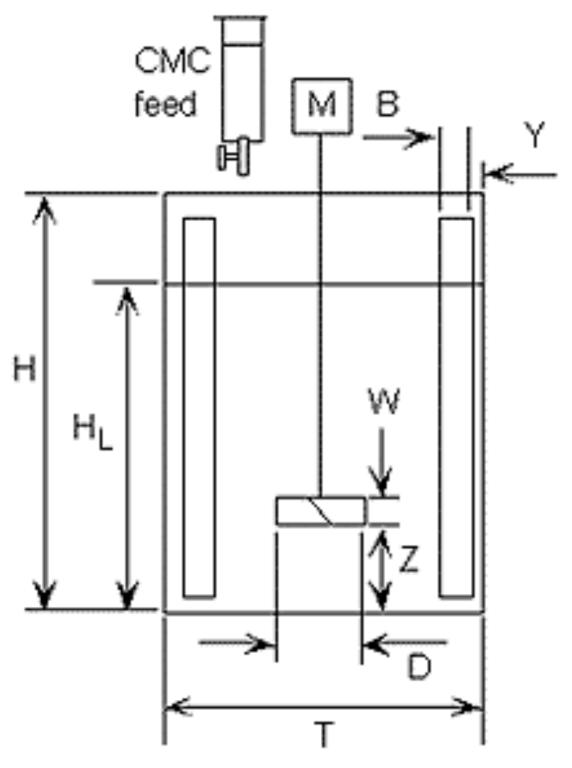
- Mass transfer
 - Nutrients
 - Oxygen
- Energy transfer
 - Heat
 - (electrons)

Type 1 MSC	θ_{I} =	$=g_{\varphi}(\varphi)$	$\theta_1 =$
Type 3 MSC	$\theta_l = g_{\varphi,T}(\varphi,T)$		$\theta_1 = 1$
		Partial deriva	tives
	$\frac{\partial \varphi}{\partial \theta}\Big _{T}$	$\frac{\partial \boldsymbol{\varphi}}{\partial T}\Big _{\boldsymbol{\theta}}$	$\left. \frac{\partial p_c}{\partial \theta} \right _T$
Equivalency	$\left. \left(\frac{\partial \theta}{\partial \varphi} \right _T \right)^{-1} = \frac{1}{\xi_{\varphi \varphi}}$	$-\frac{\partial \varphi}{\partial \theta}\Big _{T} \cdot \frac{\partial \theta}{\partial T}\Big _{\varphi}$ $= -\frac{\xi_{\varphi T}}{\xi_{\varphi \varphi}}$	$ \left. \frac{\left(\frac{\partial p_c}{\partial \varphi} \frac{\partial \varphi}{\partial \theta} \right) \right _T }{\partial \varphi} \right _T = \frac{\partial f(\varphi, T)}{\partial \varphi} \left _T \frac{1}{\xi_{\varphi\varphi}} \right _T $
Type 1 MSC	$\left(rac{dg_{arphi}}{darphi} ight)^{-1}$	0	$-\frac{R_v\rho_lT}{}$
Type 3 MSC	$\left. \left(\frac{\partial g_{\varphi,T}}{\partial \varphi} \right _T \right)^{-1}$	$-\frac{\partial g_{\varphi,T}}{\partial T}\bigg _{\varphi}\cdot \left(\frac{\partial g_{\varphi,T}}{\partial \varphi}\bigg _{T}\right)$	-1 $-\frac{1}{\varphi\xi_{\varphi\varphi}}$
		Balance equa	ntion
	$\frac{\partial \theta}{\partial t} =$	$\nabla \cdot \left[\left(D_{\theta T}^{l} + D_{\theta T}^{v} \right) \nabla T + \right.$	$\left(D_{\theta\theta}^l + D_{\theta\theta}^v\right) \nabla \theta \big]$
	Secon	dary moisture tran	sport functions
Isother	Vapor transport mal Non-	isothermal	Liqu Isothermal
	D_{ox}^{v}		222

 $\varphi \leftrightarrow \theta_1$



Geometry of standard stirred tank for aerobic reactions



Reactor Configuration				
Tank diameter	Т	105 mm		
Baffles		4 number		
Baffle width	В	T/12		
Baffle spacing	Υ	T/60		
Impeller diameter	D	T/3		
Bottom clearance	Z	T/3		
Liquid depth	Hլ	Τ		
Number of blades	n	4		
Blade width	w	D/5		
Blade angle	α	45 °		



Mycelium



Bioreactor value pyramid

- Antibiotics
- Steroids / hormones
- Vitamins
- Proteins
- Sugars
- Acids

Pharma

Food



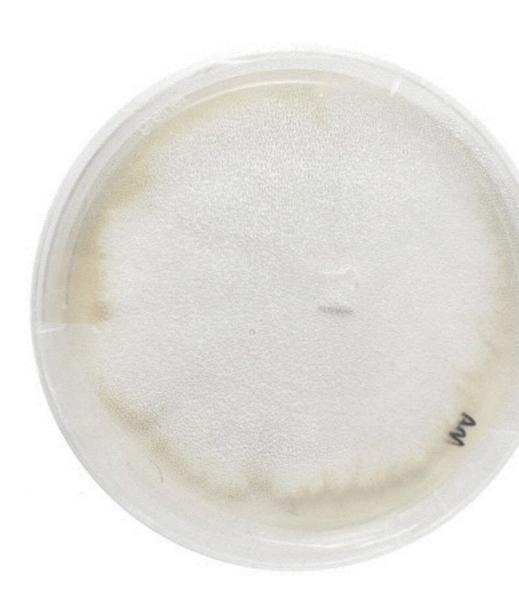
Heat

Volume



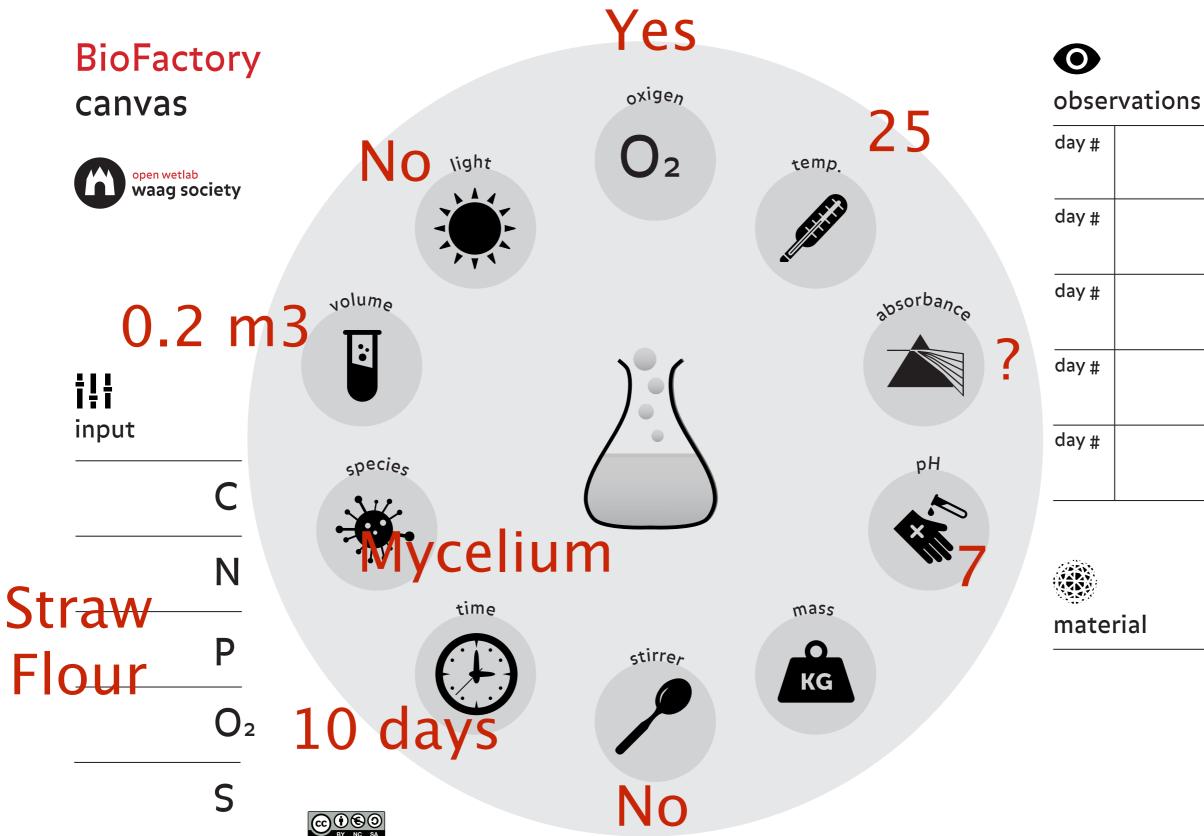
MycoMake recipe

- Straw
- Starting culture
- Water
- Flour
- Grow for 4–5 days at room temperature
 - In the dark, in an open bag
- Put the material in a mold
- Grow again for 4–5 days
 - In the dark
- Dry in an oven





Mycelium canvas





Fungal Futures





Example Production Process Design

Violacein production



Janthiobacterium lividum



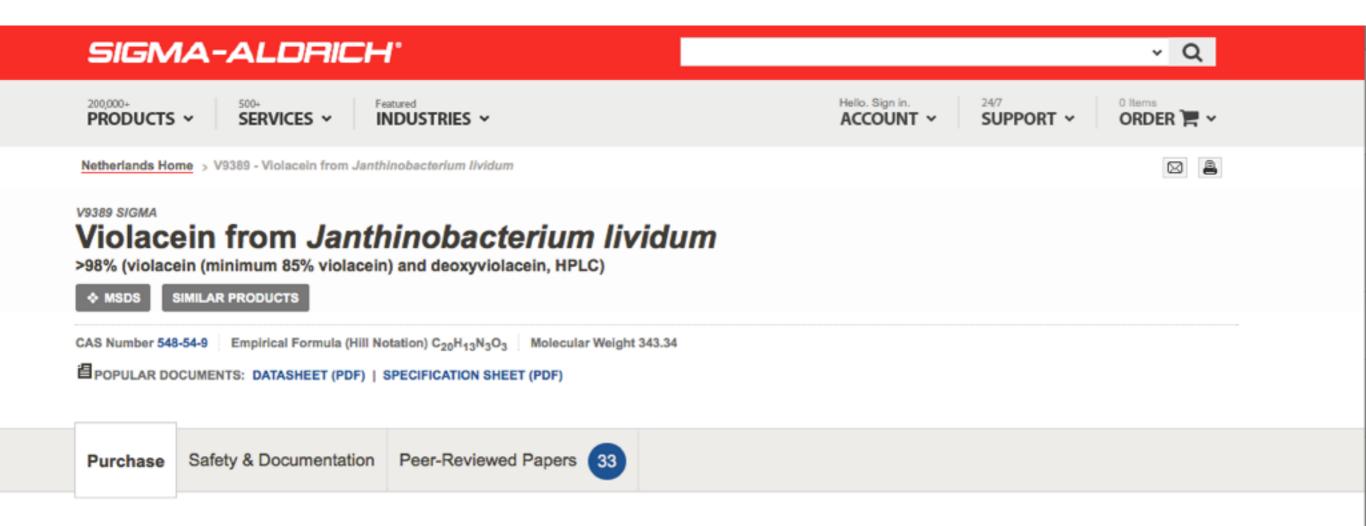


My search for J. lividum

- "Janthinobacterium lividum" +
 - "growth conditions"
 - "violacein pathway"
 - "violacein genes"
 - "patent"
 - "yield"
 - "inhibition"
 - "extraction"



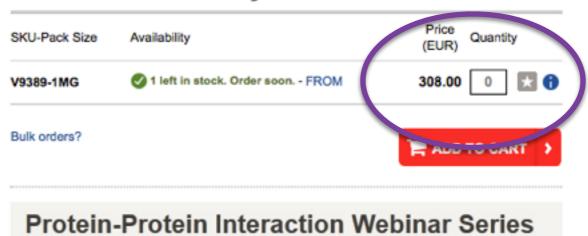




Properties

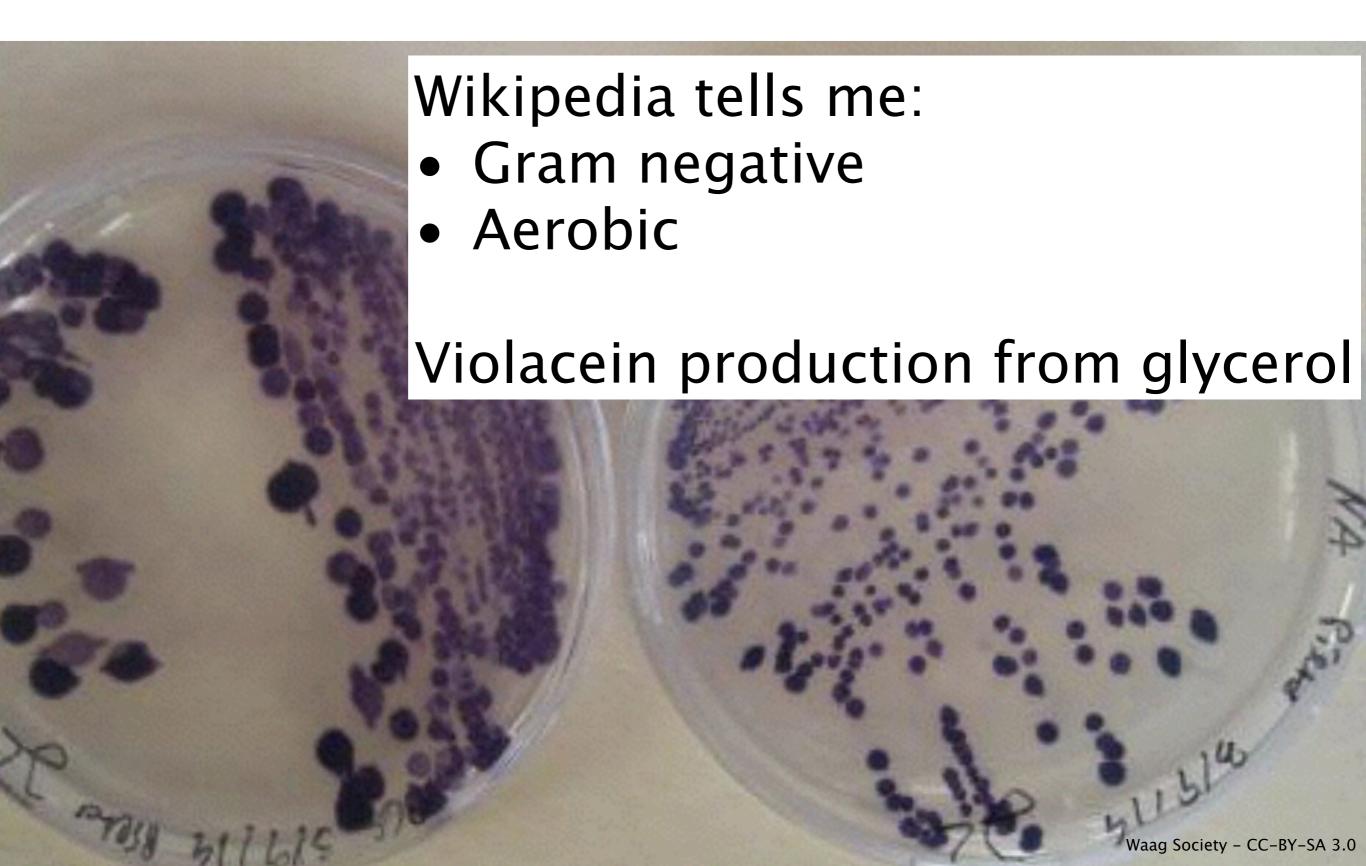
Related Categories	Apoptosis Inducers, Apoptosis and Cell Cycle, Bioactive
	Small Molecule Alphabetical Index, Bioactive Small
	Molecules, Cell Biology,
	More
assay	>98% (violacein (minimum 85% violacein) and
	deoxyviolacein, HPLC)
solubility	H ₂ O: insoluble
	acetone: soluble
	ethanol: soluble

Price and Availability



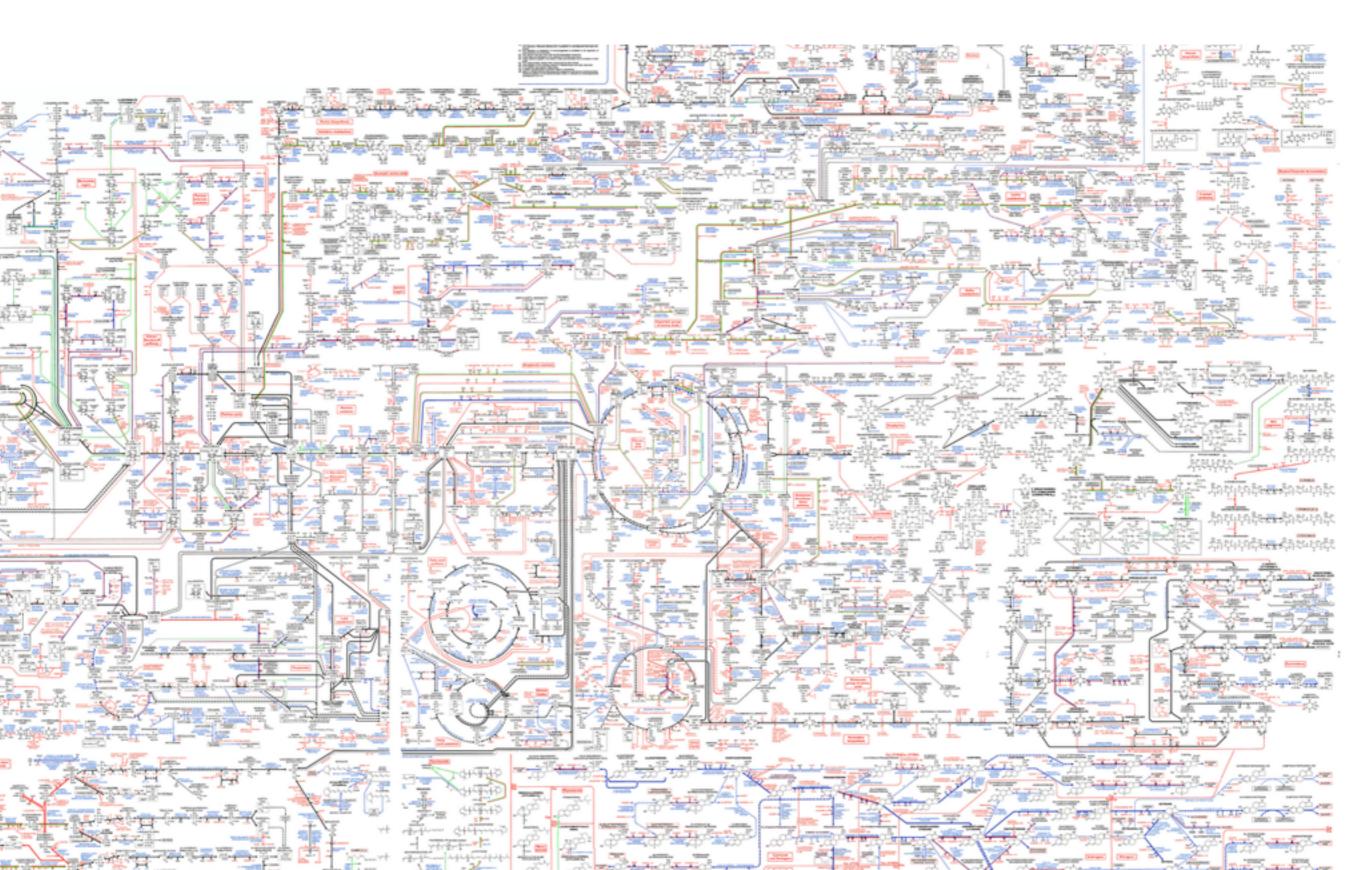


Janthinobacterium lividum





Production pathway?





P. Roqueforti eating lactate

$$C_3H_5O_3^- + 3O_2 + H^+ -> 3CO_2 + 3H_2O_3$$

Acid is consumed



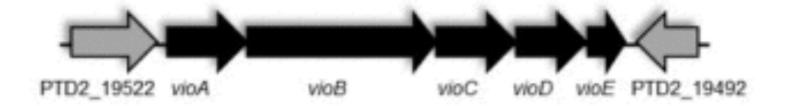
Violacein genes

Hornung et al. - The Janthinobacterium sp. HH01 Genome Encodes a Homologue of the V. cholerae CqsA and L. pneumophila LqsA Autoinducer Synthases (2013)

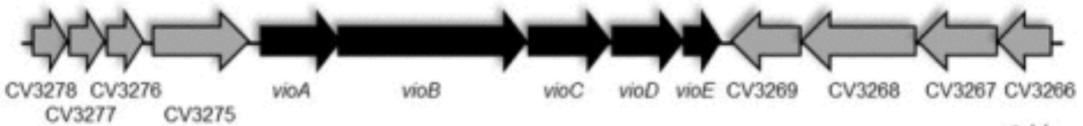
Janthinobacterium sp. HH01



Pseudoalteromonas tunicata D2



Chromobacterium violaceum ATCC 12472



Production pathway?

Tryptophan

Violacein biosynthesis in Chromobacterium violaceum COOH Tryptophan Tryptophan HOOC HO COOH 1,2 shift COOH Violacein

Figure 2. Violacein biosynthesis, as proposed by August et al., 2000. VioA, VioB, VioC, and VioD are the gene products of the biosynthesis operon, encoding nucleotide-dependent monooxygenases and a protein similar to a polyketide synthase (VioB).



Other interesting things:

• J. lividum produces a metallo- β -lactamase conferring resistance to

several β -lactam antibiotics

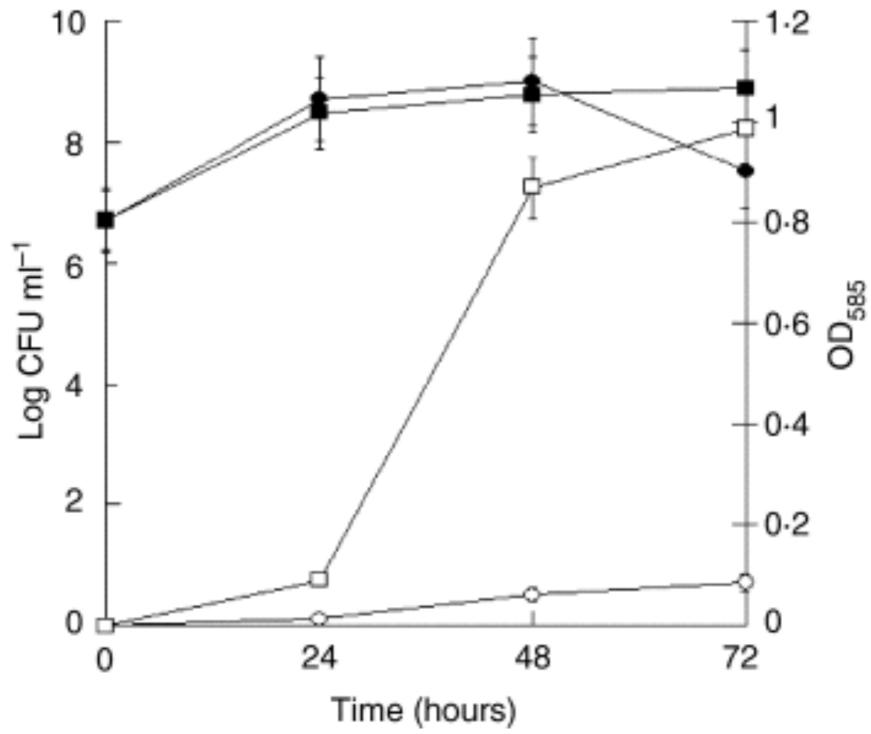
Rossolini, G.M., Condemi, M.A., Pantanella, F., Docquier, J.D., Amicosante, G. and Thaller, M.C. (2001) Metallo-β-lactamase producers in environmental microbiota: new molecular class B enzyme in Janthinobacterium lividum. Antimicrob Agents Chemother 45, 837-844.

- Violacein:
 - C₂₀-H₁₃-N₃-O₃
 - molecular weight of 343-33
 - insoluble in water
 - soluble in alcohols as methanol, ethanol and acetone
 - maximal absorption in a solution of methanol is at 585 nm

Blosser, R.S. and Gray, K.M. (2000) Extraction of violacein from Chromobacterium violaceum provides a new quantitative bioassey for N-acyl homoserine lactone autoinducers. J Microbiol Methods 40, 47-55.



Production inhibition



Pantanella, F., Berlutti, F., Passariello, C., Sarli, S., Morea, C. and Schippa, S. (2007), Violacein and biofilm production in *Janthinobacterium lividum*. Journal of Applied Microbiology, 102: 992–999. doi: 10.1111/j.1365-2672.2006.03155.x



Production conditions

Growing the bacteria in culture took 5 days before the culture would turn purple due to *J. lividum* forming a biofilm in the media. Large culture growth by embedding sterile cotton mats in sterile 2L bottles with nutrient media with the added glycerol and L-tryptophan (fig. 2) that showed purple coloring after 48 hour incubation [9]. The mats were extracted after 5 days to harvest the violacein. Yield of violacein from after crude methanol extraction and low was about 10mg.

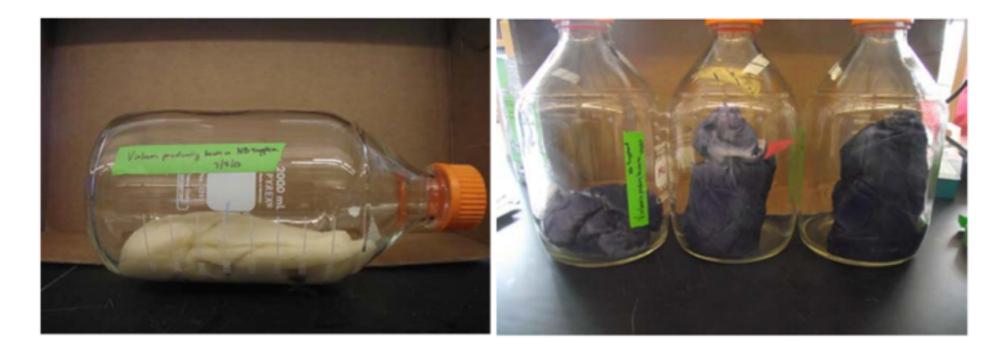


Figure 2: Violacein optimization. 1% Glycerol and 250μM L-tryptophan were added to the nutrient broth media to enhance pigment development. Cotton mats were used to allow bacteria to become sessile and produce violacein faster than with liquid cultures.



Process for the production of violacein and its derivative deoxyviolacein containing bioactive pigment from Chromobacterium sp. (MTCC5522)

EXAMPLE 1

PRODUCTION AND EXTRACTION OF THE BIOACTIVE PIGMENT FROM THE CULTURE OF CHROMOBACTERIUM SP. NIIST-CKK-01

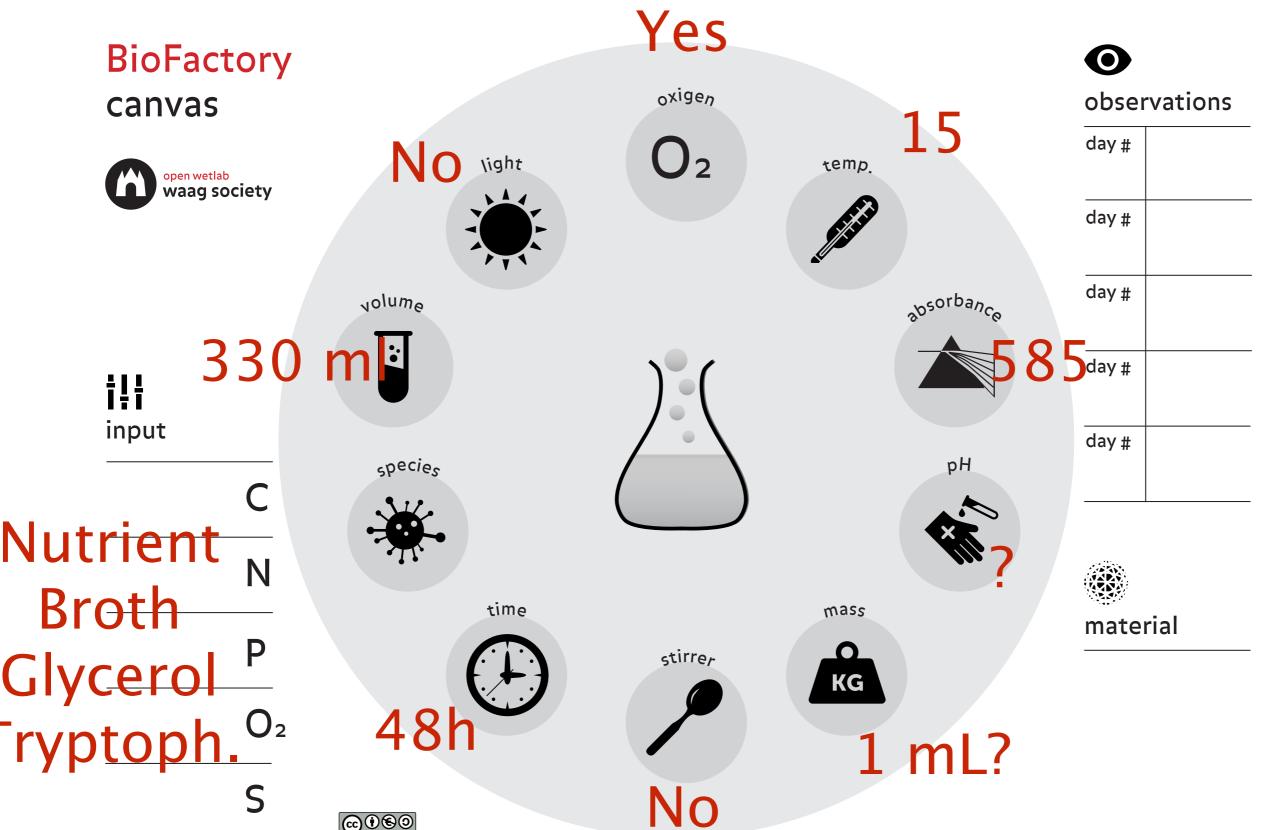
A loopful of 24 hrs old pure culture Chromobacterium sp. NIIST-CKK-01 from solid agar medium (LB agar or Nutrient agar) was inoculated with 50 ml of the growth medium (0.5% Yeast extract and 1.5% Peptone) taken in a 250 ml Erlenmeyer flask. Alternatively, 10% (v/v) of 24 hour old pure culture of Chromobacterium sp. NIIST-CKK-01 in LB broth was also used as inoculum. The pH of the medium was 7. The flasks inoculated with Chromobacterium sp. NIIST-CKK-01 were subsequently incubated in a rotary shaker at ambient temperature (30 °C) and 200 rpm for 24 hours. The deep purple purple-blue pigment starts appearing in the medium by about 6 hours of incubation and continued beyond biomass increase (Fig 1).

After 24 hrs of incubation, the bacterial biomass with pigment was centrifuged at 9676.8 x g and 4 °C for 10 minutes. After centrifugation, the clear supernatant was removed. The pellet containing biomass and pigment was mixed thoroughly with 5 ml of extra pure methanol. The mixture was centrifuged again at 9676.8 χ g and 4 °C for 10 minutes to separate the cell pellet from the solvent-pigment mixture. The pigment extraction was repeated twice using fresh solvent as described. All the pigment extracted solvent pooled together and the pigment was concentrated by normal vacuum drying in a desiccator. The quantity of biomass and pigment produced could be accounted by measuring optical density at 600 nm and 575 nm respectively. The yield of pigment by this method was about 1.0 g pigment/g of dry biomass in 24 hrs.

HPLC analysis is carried out for checking the purity of the pigment produced using an ODS column (Lichrospher-100; Merck) with acetonitrile (40%) at Iml/min as mobile phase and using UV-VIS detector at 575 nm (Figure 2). UV-VIS absorption spectra indicated maximum absorption at 575 nm, typical of violacein and its derivatives (Figure 3). EXAMPLE 2

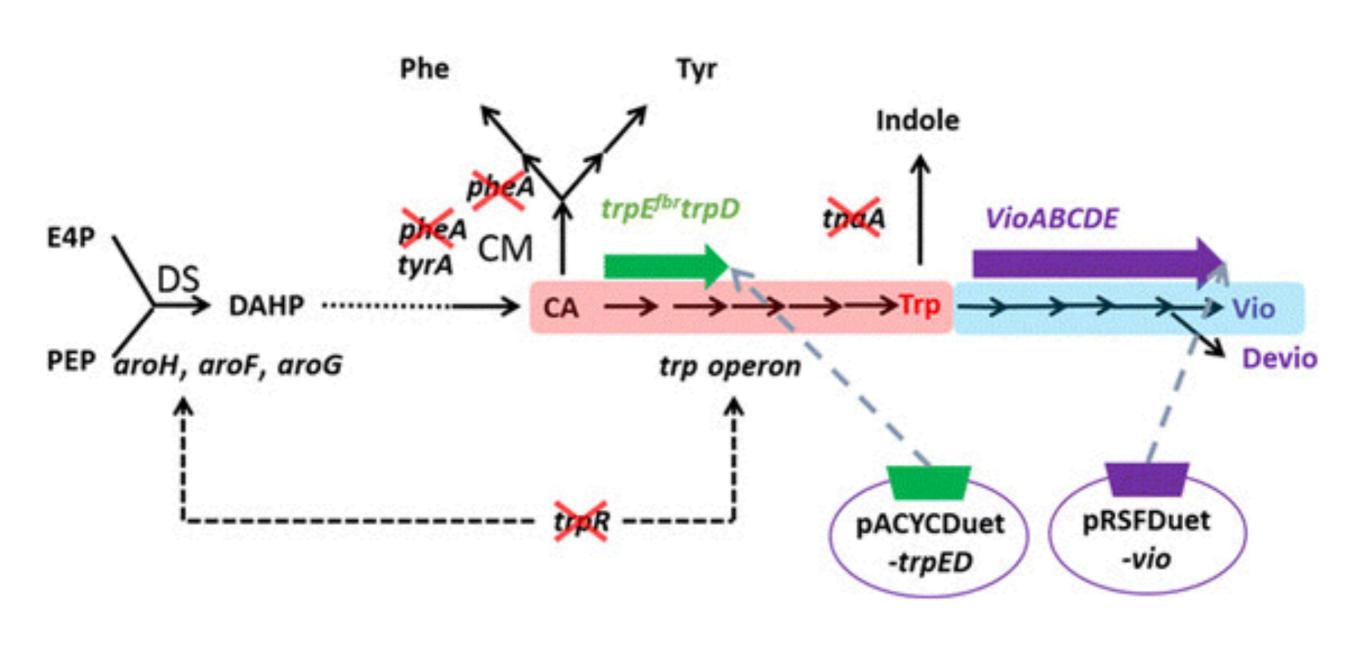


J. Lividum canvas





Genetic construct for E. coli



Fang et al. Microbial Cell Factories 2015 14:8 doi:10.1186/s12934-015-0192-x



Synbiota – ScienceHack





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4/8/14



